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HEPT DERIVATIVES: 6-BENZYL-1-ETHOXYMETHYL-5-ISOPROPYLURACIL (MKC-442)

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Abstract - The 1-[(2-hydroxyethoxy)methyl]-6-(phenylthio)thymine (HEPT) derivatives have been found to be potent and specific inhibitors of human immunodeficiency virus type 1 (HIV-1) replication *in vitro*. Among the compounds, MKC-442 (6-benzyl-1-ethoxymethyl-5-isopropyluracil) has recently been chosen as a candidate for clinical efficacy and safety studies in patients with the acquired immune deficiency syndrome (AIDS).

In the fight against human immunodeficiency virus type 1 (HIV-1), the causative agent of the acquired immune deficiency syndrome (AIDS), several classes of compounds have been identified as highly specific inhibitors of HIV-1.¹ Among the compounds, 1-[(2-hydroxyethoxy)methyl]-6-(phenylthio)thymine (HEPT) has been described in 1989 as a novel lead for specific anti-HIV-1 agents.^{2,3} After the discovery of HEPT, we have attempted at increasing its potency by introducing various chemical modifications and obtained several HEPT derivatives that inhibit HIV-1 replication in the nanomolar concentration range.^{4,5} We have also investigated their toxicologies and pharmacologies in animals to select the best candidate(s) for clinical trials. From these studies, we have chosen MKC-442 (6-benzyl-1-ethoxymethyl-5-isopropyluracil or I-EBU) as a candidate for clinical efficacy and safety studies in AIDS patients.⁶

Chemistry

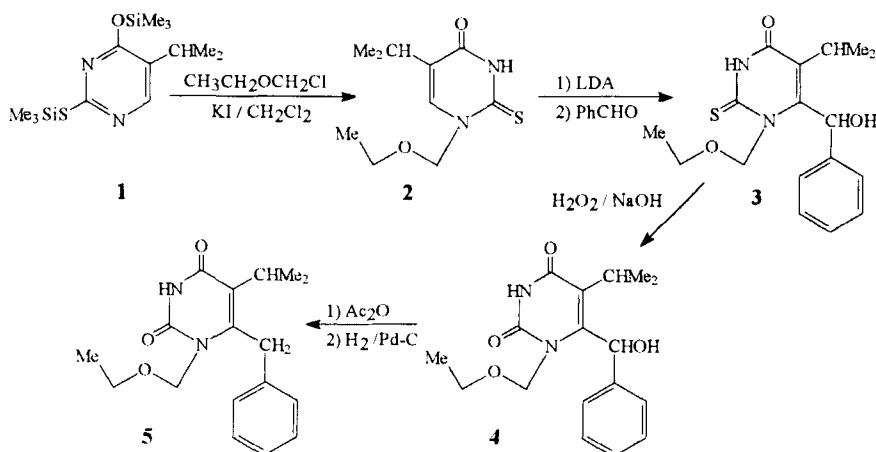
It is well-known that glycosidation of 6-substituted uracils almost invariably results in the predominant formation of N³-glycosylated products due to steric hindrance by the substituents. Therefore, the synthesis of 6-substituted uridines can be carried out best by using uridine as the starting material. We reported that regiospecific C-6 lithiation of the uracil moiety took place when 2',3'-O-isopropylideneuridine was deprotonated with lithium diisopropylamide (LDA).⁷ Since various types of electrophiles react with the C-6 lithiated species, this method disclosed a simple entry to a wide range of 6-substituted uridines, most of which are difficult to synthesize by any other methods.

HEPT was discovered during the preparation of acyclic 6-substituted uridines based on the LDA lithiation chemistry.² Although a number of acyclonucleosides had been known, no such 6-substituted pyrimidine derivatives had appeared in literature when we embarked upon the evaluation of HEPT for its antiviral activities. Extensive synthetic efforts to improve the anti-HIV-1 activity of HEPT finally led to MKC-442.

The synthesis of MKC-442 is outlined in Scheme 1. For the reasons outlined above, the 6-substituent has to be introduced after the acyclic portion has been introduced. In this particular case of MKC-442, the silylated 5-isopropyl-2-thiouracil **1** was used as the starting material, because the N¹-ethoxymethyl derivative **2** gave a much higher yield of the 6-substituted product **3** than the corresponding 5-isopropyluracil derivative did upon the LDA lithiation and subsequent reaction with benzaldehyde. Oxidative hydrolysis of **3** readily afforded **4**, which was then converted to MKC-442 by conventional hydrogenolysis.

Anti-HIV-1 activity

MKC-442 completely protected MT-4 cells against HIV-1-induced cell destruction at a concentration of 160 nM (data not shown). The 50% effective concentration (EC₅₀) of MKC-442 for the III_B strain was 15 nM (Table 1). In contrast, MKC-442 did not reduce the growth and viability of mock-infected MT-4 cells at concentrations up to 20 μM (data not shown). Its 50% cytotoxic concentration (CC₅₀) was 102 μM (Table 1). MKC-442 was also effective against HIV-1 replication in MOLT-4 cells, peripheral blood lymphocytes (PBL), and monocyte-macrophages (M/M).



Scheme 1. Synthesis of MKC-442

However, as previously noted for HEPT, MKC-442 did not inhibit HIV-2 replication in MT-4 cells (Table 1). MKC-442 was equally active against 3'-azido-3'-deoxythymidine (AZT)-resistant mutants (A012D and A018C)^{8,9} and AZT-susceptible strains (Table 1).

Inhibition of HIV-1 RT

Although the HEPT derivatives are structurally related to nucleosides, previous studies on their mechanism of action have revealed that they functionally behave as HIV-1-specific nonnucleoside RT inhibitors.^{4,5} MKC-442 also markedly inhibited HIV-1 RT activity (Table 2). When poly(C)/oligo(dG) was used as the template/primer, the 50% inhibitory concentration (IC₅₀) of MKC-442 was 0.012 μ M. However, MKC-442 was totally inactive against HIV-1 RNase H, avian myeloblastosis virus (AMV) RT, Moloney murine leukemia virus (MLV) RT, and calf thymus DNA polymerase α (Table 2). In contrast, AZTTP was inhibitory to these RTs and DNA polymerase α . The kinetic analysis revealed that the inhibition of HIV-1 RT by MKC-442 was noncompetitive, linear-mixed, and uncompetitive with respect to the substrates (dTTP and dGTP) and templates [poly(A) and poly(C)], respectively.¹⁰

Table 1. Antiviral activity of MKC-442 and AZT in cell cultures

| Virus | Strain | Cells | MKC-442 | | AZT | |
|-------|-------------------|--------|---|---|--------------------------------|--------------------------------|
| | | | EC ₅₀ ^a (μ M) | CC ₅₀ ^b (μ M) | EC ₅₀ (μ M) | CC ₅₀ (μ M) |
| HIV-1 | III _B | MT-4 | 0.015 | 102 | 0.0033 | 10 |
| | | MOLT-4 | 0.0031 | 181 | 0.00080 | 19 |
| | | PBL | 0.0038 | 95 | 0.0021 | 12 |
| | III _{RF} | MT-4 | 0.012 | - | 0.0059 | - |
| | HE | MT-4 | 0.019 | - | 0.0066 | - |
| | JR-FL | M/M | 0.011 | > 100 | 0.0038 | > 100 |
| | A012D | MT-4 | 0.0047 | - | 0.19 | - |
| | A018C | MT-4 | 0.0076 | - | 0.22 | - |
| HIV-2 | ROD | MT-4 | > 102 | - | 0.0025 | - |
| | EHO | MT-4 | > 102 | - | 0.0039 | - |

^a Except for A012D and A018C strains, the 50% effective concentration (EC₅₀) was based on the inhibition of HIV-induced cytopathic effect in MT-4 and MOLT-4 cells or the reduction of p24 antigen in culture supernatants of PBL and M/M. For A012D and A018C, the EC₅₀ was based on the inhibition of HIV-1 antigen expression in MT-4 cells.

^b The 50% cytotoxic concentration (CC₅₀) was based on the reduction of viability of mock-infected cells.

Data are taken from reference 6.

Combination with other compounds

The combined inhibitory effect of MKC-442 and AZT, 2',3'-dideoxycytidine (DDC), or 2',3'-dideoxyinosine (DDI) on HIV-1 replication was found to be synergistic in MT-4 cells.⁶ Furthermore, unlike other non nucleoside RT inhibitors, MKC-442 acted also synergistically with AZTTP at the level of the HIV-1 RT.¹⁰ In agreement with these observations,

Table 2. Effect of MKC-442 and AZT on various RTs and DNA polymerase α

| Enzyme | Template/primer | IC ₅₀ ^a (μ M) | |
|-------------------------------|-------------------|--|--------------------|
| | | MKC-442 | AZTTP ^c |
| HIV-1 RT | poly(A)/oligo(dT) | 0.21 | 0.007 |
| | poly(C)/oligo(dG) | 0.012 | > 1000 |
| HIV-1 RNaseH | - | > 1000 | > 1000 |
| AMV RT | poly(A)/oligo(dT) | > 1000 | 0.12 |
| MLV RT | poly(A)/oligo(dT) | > 1000 | 0.20 |
| DNA pol α ^b | activated DNA | > 1000 | 540 |

^a 50% Inhibitory concentration, or concentration required to inhibit enzyme activity by 50%.

^b Calf thymus DNA polymerase α .

^c AZT 5'-triphosphate.

Data are taken from reference 10.

we found that the simultaneous treatment of HIV-1-infected MT-4 cells with MKC-442 and AZT could prevent the breakthrough of virus for more than 60 days (Fig. 1).

Drug resistance

Recent studies on HIV-1 mutants resistant to nonnucleoside RT inhibitors have demonstrated that single amino acid changes in the RT are responsible for resistance.¹²⁻¹⁵ We have also obtained drug-resistant HIV-1 mutants upon several passages in cell culture in the presence of MKC-442. The mutants were cross-resistant to nevirapine¹⁶ and L-696,229,¹⁷ but not to AZT (Table 3). Sequence analysis of RT revealed that the HIV-1 mutant completely resistant to MKC-442 (HE₄₄₂-2) had two amino acid substitutions (Lys¹⁰³ \rightarrow Arg and Tyr¹⁸¹ \rightarrow Cys), whereas the mutants partially resistant to MKC-442 (III_B-R and HE₄₄₂-1) had only the Tyr¹⁸¹ \rightarrow Cys substitution.¹⁸

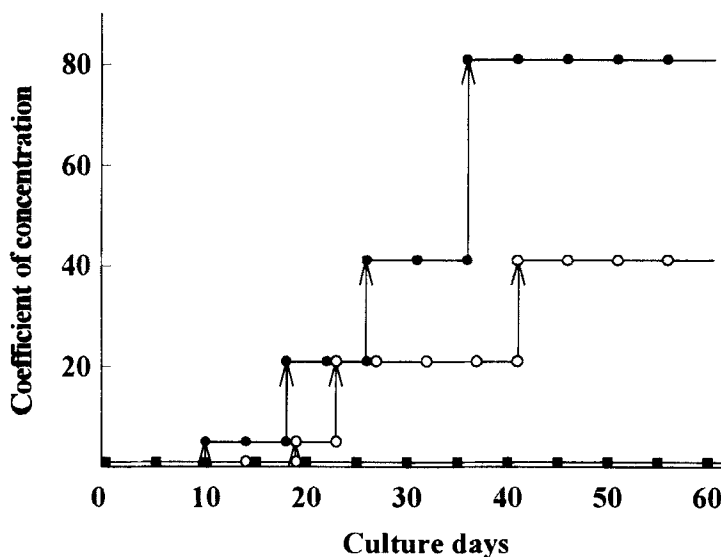


Fig. 1. Selection of drug-resistant HIV-1 in the presence of compounds. MT-4 cells were infected with III_B strain and cultured in the presence of either MKC-442 alone (●), AZT alone (○), or combination of MKC-442 and AZT (■). The starting concentrations of MKC-442 alone and AZT alone were 56 and 16 nM, respectively. For the combination, 28 and 8 nM were used as the initial concentrations of MKC-442 and AZT, respectively. After a 4- or 5-day incubation at 37°C, the cells were passed by 1:5 dilution into fresh culture medium containing the same concentration of the compound. When the number of viable cells was less than 1×10^5 per ml due to the HIV-1-induced cytopathic effect, subsequent passage was not performed. In this case, the culture supernatant was collected and examined for its infectivity. Then MT-4 cells were infected with the virus at a MOI of 0.1 and cultured in 1 ml of medium containing MKC-442 at a concentration which was 2- to 5-fold higher than that in the previous passage. The concentration of compounds in each passage was expressed as a ratio to the starting concentration (coefficient of concentration). Data are taken from reference 11.

Table 3. Antiviral activity of MKC-442, nevirapine, L-696,229, and AZT against MKC-442-resistant strains

| Strain | EC ₅₀ ^a (μM) | | | |
|---------------------|------------------------------------|------------|-----------|--------|
| | MKC-442 | Nevirapine | L-696,229 | AZT |
| IIIB | 0.015 | 0.077 | 0.022 | 0.0033 |
| IIIB-R | 53 | > 224 | > 188 | 0.0023 |
| HE | 0.016 | 0.073 | 0.031 | 0.0060 |
| HE ₄₄₂₋₁ | 88 | > 191 | > 179 | 0.0026 |
| HE ₄₄₂₋₂ | > 119 | > 191 | > 179 | 0.0047 |

^a The 50% effective concentration (EC₅₀) was based on the inhibition of HIV-1-induced cytopathic effect in MT-4 cells.

Data are taken from references 6 and 18.

Table 4. Antiviral activity of MKC-442, nevirapine, L-696,229, and AZT against A018A, A018C, and A018CR

| Compound | EC ₅₀ ^a (μM) | | |
|------------|------------------------------------|--------|--------|
| | A018A | A018C | A018CR |
| AZT | 0.0015 | 0.31 | 0.070 |
| DDI | 11 | 11 | 6.8 |
| MKC-442 | 0.0059 | 0.0088 | 1.5 |
| Nevirapine | 0.030 | 0.043 | 0.074 |
| L-696,229 | 0.027 | 0.036 | 0.28 |

^a The 50% effective concentration (EC₅₀) was based on the inhibition of HIV-1 antigen expression in MT-4 cells.

Data are taken from reference 11.

The HIV-1 mutant co-resistant to MKC-442 and AZT could be obtained in cell culture when the selection experiment was started with the AZT-resistant strain A018C. Table 4 shows the inhibitory effects of various HIV-1 RT inhibitors on the replication of A018A (AZT-susceptible), A018C (AZT-resistant), and A018CR (co-resistant to MKC-442 and AZT) in MT-4 cells. A018CR was found to be highly resistant to MKC-442. Interestingly, A018CR retained almost normal sensitivity to nevirapine; it was also more susceptible to AZT than was A018C, although it was less AZT-susceptible than A018A. In addition to the amino acid substitutions that have been reported to confer AZT-resistance,⁹ A018CR showed the substitutions Leu⁷⁴ → Ile, Lys¹⁰³ → Glu, and Val¹⁰⁸ → Ile.¹¹ It remains to be elucidated whether the combination chemotherapy will achieve prolonged suppression of drug-resistant mutants.

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REFERENCES

1. E. De Clercq, *Med. Res. Rev.* 13, 229-258 (1993).
2. T. Miyasaka, H. Tanaka, M. Baba, H. Hayakawa, R.T. Walker, J. Balzarini and E. De Clercq, *J. Med. Chem.* 32, 2507-2509 (1989).
3. M. Baba, H. Tanaka, E. De Clercq, R. Pauwels, J. Balzarini, D. Schols, H. Nakashima, C.-F. Perno, R.T. Walker and T. Miyasaka, *Biochem. Biophys. Res. Commun.* 165, 1375-1381 (1989).
4. M. Baba, E. De Clercq, H. Tanaka, M. Ubasawa, H. Takashima, K. Sekiya, I. Nitta, K. Umezu, H. Nakashima, S. Mori, S. Shigeta, R.T. Walker and T. Miyasaka, *Proc. Natl. Acad. Sci. USA* 88, 2356-2360 (1991).
5. M. Baba, E. De Clercq, H. Tanaka, M. Ubasawa, H. Takashima, K. Sekiya, I. Nitta, K. Umezu, R.T. Walker, S. Mori, M. Ito, S. Shigeta and T. Miyasaka, *Mol. Pharmacol.* 39, 805-810 (1991).
6. M. Baba, S. Shigeta, S. Yuasa, H. Takashima, K. Sekiya, M. Ubasawa, H. Tanaka, T. Miyasaka, R.T. Walker and E. De Clercq, *Antimicrob. Agents Chemother.* 38, 688-692 (1994).

7. H. Tanaka, H. Hayakawa and T. Miyasaka, *Tetrahedron* 38, 2635-2642 (1982).
8. B.A. Larder, G. Darby and D.D. Richman, *Science* 243, 1731-1734 (1989).
9. B.A. Larder and S.D. Kemp, *Science* 246, 1155-1158 (1989).
10. S. Yuasa, Y. Sadakata, H. Takashima, K. Sekiya, N. Inouye, M. Ubasawa and M. Baba, *Mol. Pharmacol.* 44, 895-900 (1993).
11. M. Baba, K. Hashimoto, S. Shigeta, M. Seki, Y. Sadakata and S. Yuasa, Manuscript submitted for publication (1994).
12. D. Richman, C.-K. Shih, I. Lowy, J. Rose, P. Prodanovich, S. Goff and J. Griffin, *Proc. Natl. Acad. Sci. USA* 88, 11241-11245 (1991).
13. J.H. Nunberg, W.A. Schleif, E.J. Boots, J.A. O'Brien, J.C. Quintero, J.M. Hoffman and E.A. Emini, *J. Virol.* 65, 4887-4892 (1991).
14. J. Balzarini, A. Karlsson, M.-J. Pérez-Pérez, L. Vrang, J. Walbers, H. Zhang, B. Oberg, A.-M. Vandamme, M.-J. Camarasa and E. De Clercq, *Virology* 192, 246-253 (1993).
15. E. De Clercq, *Biochem. Pharmacol.* 47, 155-169 (1994).
16. V.J. Merluzzi, K.D. Hargrave, M. Labadia, K. Grozinger, M. Skoog, J.C. Wu, C.-K. Shih, K. Eckner, S. Hattox, J. Adams, A.S. Rosethal, R. Faanes, R.J. Eckner, R.A. Koup and J.L. Sullivan, *Science* 250, 1411-1413 (1990).
17. M.E. Goldman, J.A. O'Brien, T.L. Ruffing, J.H. Nunberg, W.A. Schleif, J.C. Quintero, P.K.S. Siegl, M.J. Hoffman, A.M. Smith and E.A. Emini, *Antimicrob. Agents Chemother.* 36, 1019-1023 (1992).
18. M. Seki, Y. Sadakata, S. Yuasa and M. Baba, Manuscript submitted for publication (1994).